

- 1 To whom reprints should be addressed.
- 2 Katz, A.M., and Messineo, F.C., *Circulation Res.* 48 (1981) 1.
- 3 Dhalla, N.S., Das, P.K., and Sharma, G.P., *J. molec. cell. Cardiol.* 10 (1978) 363.
- 4 Papahadjopoulos, D., Cowden, M., and Kimelberg, H., *Biochim. biophys. Acta* 330 (1973) 8.
- 5 Hasin, J., Shimoui, J., Stein, O., and Stein, J., *J. molec. cell. Cardiol.* 12 (1980) 675.
- 6 Lane, L.K., Anner, B.M., Wallick, E.T., Ray, M.V., and Schwartz, A., *Biochem. Pharmacol.* 27 (1978) 225.
- 7 Morrison, E.S., Scott, R.F., Imai, H., Kroms, M., Nour, B.A., and Briggs, R.G., *Atherosclerosis* 12 (1970) 139.
- 8 Günther, J., Kutscherskij, E., Storch, E., and Vetter, R., *Acta biol. med. germ.* 41 (1982) 325.
- 9 Reuter, H., *Circulation Res.* 34 (1974) 599.
- 10 Harris, P., *Eur. J. Cardiol.* 3/2 (1975) 157.
- 11 Lamers, J.M.J., Stinis, J.T., Kort, W.J., and Hülsmann, W.C., *J. molec. cell. Cardiol.* 10 (1978) 235.
- 12 Gornall, A.G., Bardawill, Ch.J., and David, M.M., *J. biol. Chem.* 177 (1949) 751.
- 13 Bondke, H., Grosse, J., Lemke, B., Ott, G., and Pohle, M., Thesis Humboldt-Universität, Berlin 1979.
- 14 Crane, F.L., Glenn, J.L., and Green, D.E., *Biochim. biophys. Acta* 22 (1956) 475.
- 15 Folch, J., Lees, M., and Sloane Stanley, G.H., *J. biol. Chem.* 226 (1957) 497.
- 16 Zlatkis, A., Zak, B., and Boyle, A.J., *J. Lab. clin. Med.* 41 (1953) 486.
- 17 Rouser, G., Fleischer, S., and Jamamoto, A., *Lipids* 5 (1970) 494.
- 18 Hallermayer, G., und Neupert, W., *Hoppe-Seyler's Z. physiol. Chem.* 355 (1974) 279.
- 19 Lowry, O.H., Rosebrough, N.J., Farr, A.L., and Randall, R.J., *J. biol. Chem.* 193 (1951) 265.
- 20 Kutscherskij, E., Thesis, Humboldt-Universität, Berlin 1981.
- 21 Kutscherskij, E., and Günther, J., *J. molec. cell. Cardiol.* 12 suppl. 1 (1980) 87.
- 22 Innis, S.M., and Clandinin, M.T., *Biochem. J.* 198 (1981) 231.
- 23 Dewailly, P., Nouvelot, H., Sezille, G., Fruchart, J.C., and Jaillard, J., *Lipids* 13 (1978) 301.
- 24 Schwartz, A., Lindenmayer, G.E., and Allen, J.C., *Pharmac. Rev.* 27 (1975) 3.
- 25 Langer, G.A., *A. Rev. Physiol.* 44 (1982) 435.
- 26 Liu, M.-S., and Onji, T., *J. molec. cell. Cardiol.* 12 (1980) 1427.
- 27 Askari, A., and Rao, S.N., *Biochim. biophys. Acta* 241 (1971) 74.
- 28 Akera, T., Jamamoto, S., Chubb, J., McNish, R., and Brody, T.M., *Naunyn-Schmiedeberg's Arch. Pharmacol.* 308 (1979) 81.
- 29 Mitschelen, J.J., Clair, R.W.S., and Hester, S.H., *Arteriosclerosis* 1 (1981) 134.
- 30 Severs, N.J., *J. cell. Biol.* 25 (1981) 289.
- 31 Naito, M., *Cell Struct. Funct.* 3 (1978) 219.
- 32 Kimelberg, H.K., *Biochim. biophys. Acta* 249 (1975) 647.
- 33 Giraud, F., Claret, M., Bruckdorfer, K.R., and Chailley, B., *Biochim. biophys. Acta* 647 (1981) 249.
- 34 Bonn, F., and Greeff, K., *Archs int. Pharmacodyn. Théor.* 233 (1978) 53.
- 35 Hoerter, J.A., and Vassort, G., *J. molec. cell. Cardiol.* 12 (1980) suppl. 1, 58.
- 36 David, H., Wilfert, K., Pfeiffer, C., Günther, J., Gross, V., und Behrisch, D., *Acta biol. med. germ.* 37 (1978) 577.

0014-4754/84/080812-04\$1.50 + 0.20/0
© Birkhäuser Verlag Basel, 1984

Release of labile cyclo-oxygenase products of arachidonic acid from kidney by endotoxin¹

K. Özsan, V. İcöz and R.K. Türker

Departments of Microbiology, Thoracic Surgery and Pharmacology, Faculty of Medicine, University of Ankara, Ankara (Turkey), 16 May 1983

Summary. The possible release of prostaglandin (PG)-like substances was studied in isolated perfused kidneys from intact and from intrarenal endotoxin (Lipopolysaccharide-LPS)-injected rabbits, using the venous outflow superfusion assay organ technique. Injection of LPS into the renal artery of an LPS-pretreated kidney caused a release of thromboxane A₂ (TXA₂) and prostacyclin (PGI₂)-like materials into the venous effluent as verified by the responses of the specific assay organs. No detectable release of these substances was found in the venous outflow of LPS-injected intact kidney. The possible role of labile cyclo-oxygenase products of arachidonic acid in the Shwartzman reaction is discussed.

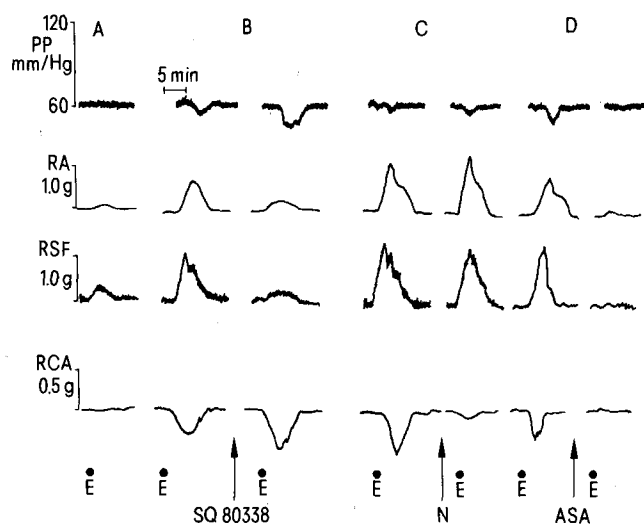
It has been shown that many hemodynamic events induced by LPS are accompanied by an increased release of PGs from the lung². This observation led to the assumption that PGs might be involved in pathological derangements of the tissues by LPS. The results of a recent study indicate that LPS may cause the release of PGs from the lung in vivo but not in vitro conditions³. It has been shown that kidney contains stable and unstable PGs and this organ can readily metabolize these lipids⁴. The present study was undertaken to investigate the possible action of LPS on the release of PGs from isolated perfused intact and LPS-pretreated rabbit kidney.

Material and methods. The experiments were carried out on adult rabbits of either sex weighing 2.0–3.0 kg. The animals were anesthetized with sodium pentobarbital (35 mg/kg i.v.) and the left kidneys were isolated, perfused with warmed (37°C) and oxygenated (5% CO₂ in O₂) Krebs' solution as described previously⁵. A group of rabbits were prepared for LPS treatment. For this procedure the animals were anesthetized with sodium pentobarbital and after the anesthesia was established an incision was made on the left flank and the kidney

was exposed. LPS (*E. coli*, lipopolysaccharide W.0111:B₄, Difco Laboratories) was injected intracortically in the upper pool of the kidney at the dose of 100 µg in 0.1 ml saline. In a group of animals (4 rabbits) only saline (0.1 ml) was injected intracortically. After that the flank was surgically sutured and the animals were kept in room temperature in separate rabbit cages. They were allowed food freely, and ordinary water. After 24 h the LPS-injected kidneys were isolated, perfused with Krebs' solution. The venous return was continuously superfused over a series of assay organs prepared in cascade⁶. The contamination of the venous outflow by urine was eliminated by a polyethylene cannula inserted into the ureter. Spirally cut rabbit aorta (RA)⁷, rat stomach fundus (RSF)⁸ and rabbit coeliac artery (CA)⁹ were used as assay organ. These assay organs were selected for the separation of TXA₂ and PGI₂-like material in the kidney outflow. TXA₂ consistently produced a contractile response in 3 assay organs^{4,9} while PGI₂ a definite relaxation in CA with extremely low concentrations⁹. The initial tension applied was 0.5–1.0 g, and the contractions were recorded on a Grass polygraph (Model 79

D) through force-displacement transducers (Grass FT-03). Perfusion pressure (PP) of the kidney was also recorded by a Statham pressure transducer (P 23 Dc). Kidneys were allowed to perfuse with normal Krebs' solution and the assay organs were superfused with the venous outflow of the kidney for a 1-h equilibration period, then LPS was applied through the renal artery by bolus injections. Aspirin (lysine acetylsalicylic acid: ASA, Bayer, Germany) as an inhibitor of PG-biosynthesis¹⁰, nicotine (nicotine tartrate:N, Geigy, Switzerland) as an inhibitor of PGI₂-biosynthesis¹¹ and SQ 80338 (Squibb, USA) as an inhibitor of TXA₂ synthesis¹² were added to the perfusion medium, depending on the experimental procedure. Sodium salt of PGI₂ (Upjohn, USA) was dissolved in 0.1 M Na₂CO₃ at the concentration of 10⁻⁴M (pH:11) and further dilutions were made in Krebs' solution immediately before use.

Results and discussion. LPS, when injected through the renal artery of intact kidney at the concentrations of 50–100 µg/ml, did not induce a response in PP but caused a slight contraction in RA and RSF. Direct superfusion of LPS over the assay organs at the same concentrations produced almost the same responses. These findings indicate that LPS did not cause a measurable release of active substance(s) from the kidney, supporting the results obtained in isolated lung tissue³. LPS (50–100 µg/ml), however, when tested in LPS-treated kidney produced a fall in PP, a contraction in venous outflow superfused RA and RSF and a relaxation in CA. The contraction of RA and RSF could partly be prevented by SQ 80338 (10⁻⁶M) but decrease in PP and relaxation of CA slightly potentiated. Inhibition by SQ 80338, a specific inhibitor of TXA₂-synthetase¹²,



Recorder tracings from isolated perfused rabbit kidney and venous effluent superfused rabbit aorta (RA), rat stomach fundus (RSF) and rabbit coeliac artery (CA) prepared in cascade. Perfusion pressure (PP) was also recorded simultaneously. Column A shows the effect of endotoxin (E) injected into the renal artery of an intact kidney. Slight contraction was observed in RA and RSF. Direct superfusion of endotoxin over the assay organs produced similar responses indicating that no appreciable release of active material occurred in intact kidney on treatment with endotoxin. Columns B, C, D represents the effect of endotoxin before and after SQ 80338 (10⁻⁶ M), nicotine: N (10⁻⁶ M) and aspirin: ASA (10⁻⁵ M) in different endotoxin-injected rabbit kidneys. Inhibition by SQ 80338, a TXA₂-synthetase inhibitor, of the contractile responses of RA and RSF, and potentiation of the relaxation observed in CA and renal PP indicate the presence of TXA₂ and PGI₂ in the venous return. Prevention by N of the relaxation observed in CA indicates the release of PGI₂. Prevention by ASA of all responses again indicates the release of labile cyclo-oxygenase products (TXA₂ and PGI₂) in the venous outflow of endotoxin-injected perfused kidney by endotoxin. Endotoxin was injected into the renal artery at a dose of 50 µg.

of the responses in RA and RSF suggests that LPS activates the TXA₂ pathway in LPS treated kidney. The increase in the fall of PP and in the relaxation of CA following SQ 80338 support this speculation, since more substrate would be available for the production of PGI₂. N, however, as specific inhibitor of PGI₂-synthetase¹¹, significantly prevented the relaxation induced by LPS without altering the responses obtained in PP, RA and RSF. ASA (10⁻⁵M), as an inhibitor of cyclo-oxygenase, when added to the perfusion medium completely abolished the responses obtained in all parameters (fig.). However, bolus injection of PGI₂ into the renal artery produced a long-lasting fall in PP and a concentration-dependent relaxation in CA. Depending on the response of CA, the possible release of PGI₂-like substance by LPS was calculated and found to be 80.0 ± 12.0 pg/ml/5 min (n:28) while no detectable release of PGI₂-like substance was found in intact kidney. These results indicate that LPS can cause the release of labile PGs from isolated perfused kidney pretreated with LPS. The production of generalized Schwartzman reaction is observed in the rabbits after 2 i.v. injections of LPS given 20–22 h apart¹³. A similar generalized Schwartzman reaction is also observed following a 2nd i.v. injection of LPS in the rabbits which were given LPS into the renal cortex beforehand. Light microscope studies have shown local necrosis, leucocytic infiltration and vascular dilatation in LPS-injected kidney (in preparation for publication). Thus, it is assumed that prior injection of LPS into the renal cortex may prepare a basis for the release of PGs (mainly TXA₂ and PGI₂) for the 2nd injection of LPS into the renal artery. The Schwartzman reaction was first described as a local skin lesion basically due to the LPS of an injected bacterial culture¹⁴. This reaction has also been observed in rabbit colon¹⁵ and heart¹⁶. Several authors have presented evidence that kinin-peptides, serotonin may have some role in the production of this reaction^{16,17,18}. The results presented here suggest that labile cyclo-oxygenase metabolites of arachidonic acid released from LPS pretreated kidney by LPS may participate in the production of generalized Schwartzman-like phenomena.

- 1 The authors are indebted to Upjohn, Kalamazoo (USA) for the generous gift of PGI₂ and to Squibb, New Jersey (USA) for SQ 80338.
- 2 Anderson, F.L., Tsagaris, I.J., Jubis, W., and Kuida, H., *Am. J. Physiol.* 228 (1975) 1479.
- 3 Feverstein, N., and Ramwell, P.W., *Br. J. Pharmac.* 73 (1981) 511.
- 4 Mc Giff, J.C., *A. Rev. Pharmac. Toxic.* 21 (1981) 479.
- 5 Türker, R.K., and Ercan, Z.S., *Prostaglandins* 9 (1975) 695.
- 6 Vane, J.R., *Br. J. Pharmac.* 23 (1964) 360.
- 7 Furchgott, R.F., and Bhadrakom, S., *J. Pharmac. exp. Ther.* 108 (1953) 129.
- 8 Vane, J.R., *Br. J. Pharmac.* 12 (1957) 344.
- 9 Bunting, S., Gryglewski, R., Moncada, S., and Vane, J.R., *Prostaglandins* 12 (1976) 897.
- 10 Vane, J.R., *Nature New Biol.* 231 (1971) 232.
- 11 Wennmalm, A., *Br. J. Pharmac.* 69 (1980) 545.
- 12 Greenberg, R., Antonaccio, M.J., and Steinbacker, T., *Eur. J. Pharmac.* 80 (1982) 19.
- 13 Hawes, E. Jr, and Mc Kay, D.G., *Haemostasis* 6 (1977) 244.
- 14 Schwartzman, G., *J. exp. Biol.* 48 (1928) 247.
- 15 Patterson, M., Terrel, C.C., O'Bryan, B.C., and Waldron, R.L., *Text. Biol. Med.* 20 (1962) 658.
- 16 Özsan, K., Türker, R.K., Özkan, U., and İçöz, V., *Ärzt. Forsch.* 25 (1975) 564.
- 17 Antapol, W., and Chrysanthos, C., *Archs Path.* 78 (1964) 313.
- 18 Chrysanthos, C., and Antapol, W., *Proc. Soc. exp. Biol. Med.* 108 (1961) 587.